

 **My select** sampler set

## Anti-RFP mAb

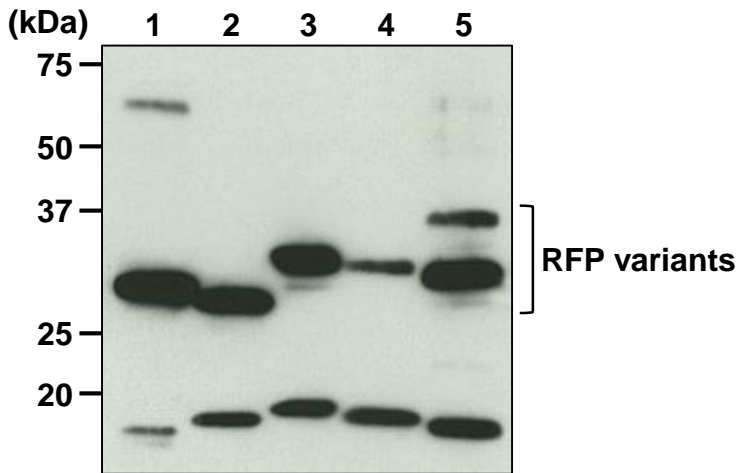
<b>CODE No.</b>	M204-3MS
<b>CLONALITY</b>	Monoclonal
<b>CLONE</b>	1G9
<b>ISOTYPE</b>	Mouse IgG2b $\kappa$
<b>QUANTITY</b>	20 $\mu$ L, 1 mg/mL
<b>SOURCE</b>	Purified IgG from hybridoma supernatant
<b>IMMUNOGEN</b>	RFP recombinant protein
<b>REACTIVITY</b>	This clone reacts with mRFP1, DsRed, mCherry, mOrange and mPlumn. It does not cross-react with GFP.
<b>FORMULATION</b>	PBS containing 50% glycerol (pH 7.2). No preservative is contained.
<b>STORAGE</b>	This antibody solution is stable for one year from the date of purchase when stored at -20°C.
<b>APPLICATION-CONFIRMED</b>	
<u>Western blotting</u>	1 $\mu$ g/mL
<u>Immunoprecipitation</u>	Not recommended
<u>Immunocytochemistry</u>	Not recommended

For more information, please visit our web site <https://ruo.mbl.co.jp/>.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

### **SDS-PAGE & Western blotting**

- 1) Mix the sample with equal volume of Laemmili's sample buffer.
- 2) Boil the sample for 3 min. and centrifuge. Load 10  $\mu$ L of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel (12.5% acrylamide) for electrophoresis.
- 3) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm<sup>2</sup> for 1 hr. in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% methanol). See the manufacturer's manual for precise transfer procedure.
- 4) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) overnight at 4°C.
- 5) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 min. x 3).
- 6) Incubate the membrane with primary antibody diluted with 1% skimmed milk (in PBS, pH 7.2) as suggested in the **APPLICATIONS** for 1 hr. at room temperature. (The concentration of antibody will depend on the conditions.)
- 7) Wash the membrane with PBS-T (5 min. x 3).
- 8) Incubate the membrane with the 1:10,000 of anti-IgG (Mouse) pAb-HRP (MBL; code no. 330) diluted with 1% skimmed milk (in PBS, pH 7.2) for 1 hr. at room temperature.
- 9) Wash the membrane with PBS-T (5 min. x 3).
- 10) Wipe excess buffer on the membrane, and then incubate it with appropriate chemiluminescence reagent for 1 min. Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.
- 11) Expose to an X-ray film in a dark room for 3 min. Develop the film as usual. The condition for exposure and development may vary.

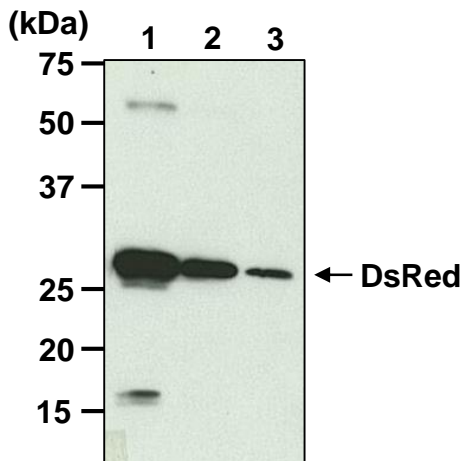


#### **Western blotting analysis of RFP variants**

- Lane 1: DsRed
- Lane 2: mRFP1\*
- Lane 3: mCherry\*
- Lane 4: mOrange\*
- Lane 5: mPlumn\*

Immunoblotted with Anti-RFP mAb (M204-3)

\*Samples were provided by RIKEN.



#### **Western blotting analysis of DsRed recombinant protein**

- Lane 1: 10 ng/lane
- Lane 2: 2 ng/lane
- Lane 3: 0.4 ng/lane

Immunoblotted with Anti-RFP mAb (M204-3)