

MONOCLONAL ANTIBODY

# FITC labeled Mouse CD11c

Code No.	Clone	Subclass	Quantity	Concentration
M100-4	223H7	Rat IgG2a	100 µL	500 µg/mL

**BACKGROUND:** The CD11c ( $\alpha X$  integrin; ~150 kDa) glycoprotein non-covalently associates with CD18 ( $\beta 2$  integrin; ~95 kDa) to form the heterodimeric complement receptor type 4 (CR4), which is involved in monocyte/granulocyte adhesion during inflammatory responses. The CD11c/CD18 receptor binds to CD54, iC3b and fibrinogen and plays a role in leukocyte adhesive interactions. CD11c/CD18 is also implicated in B cell proliferation and mediates B cell binding to fibrinogen. CD11c is commonly used as a marker for dendritic cells, but it is also expressed on macrophages, monocytes, granulocytes, NK cells, activated T and B lymphocytes and microglia.

**SOURCE:** This antibody was purified from hybridoma (clone 223H7) supernatant using protein G agarose. This hybridoma was established by fusion of mouse myeloma cell Sp2/0-Ag14 with Wister rat lymphocyte immunized with murine dendritic cells isolated from C57BL/6 mice.

**FORMULATION:** 50 µg IgG in 100 µL volume of PBS containing 1% BSA and 0.09%  $\text{NaN}_3$ .

\*Azide may react with copper or lead in plumbing system to form explosive metal azides. Therefore, always flush plenty of water when disposing materials containing azide into drain.

**STORAGE:** This antibody solution is stable for one year from the date of purchase when stored at 4°C.

**REACTIVITY:** This antibody reacts with mouse CD11c antigen on Flow cytometry.

### APPLICATIONS:

Flow cytometry: 10 µg/mL (final concentration)

\*Please refer to the data sheet (MBL code no. M100-3) for other applications.

Detailed procedure is provided in the following **PROTOCOL**.

### SPECIES CROSS REACTIVITY:

Species	Human	Mouse	Rat
Cell	Not Tested	splenocyte	Not Tested
Reactivity on FCM		+	

### INTENDED USE:

For Research Use Only. Not for use in diagnostic procedures.

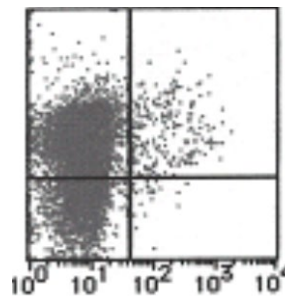
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### REFERENCES:

- 1) Fancke, B., *et al.*, *Blood* Oct 2007; 10.1182/blood-2007-05-089292.
- 2) Kruger, T., *et al.*, *J. Am. Soc. Nephrol.* **15**, 613-21 (2004)

Clone 223H7 is used in reference number 1).



**Flow cytometric analysis of Mouse CD11c expression on Mouse splenocytes.** The staining intensity of M100-4 is shown in the horizontal axis with Mouse I-A<sup>b</sup> staining on the vertical axis.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

### PROTOCOL:

#### Flow cytometric analysis for floating cells

We usually use Fisher tubes or equivalents as reaction tubes for all step described below.

- 1) Wash the cells 3 times with washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.1%  $\text{NaN}_3$ ].
- 2) Resuspend the cells with washing buffer ( $5 \times 10^6$  cells/mL).
- 3) Add 50 µL of the cell suspension into each tube, and centrifuge at 500 x g for 1 minute at room temperature (20~25°C). Remove supernatant by careful aspiration.
- 4) Add 20 µL of normal goat serum containing 1 mg/mL normal human IgG and 0.1%  $\text{NaN}_3$  to the cell pellet after tapping. Mix well and incubate for 5 minutes at room temperature.

- 5) Add 40  $\mu$ L of the FITC labeled mouse CD11c monoclonal antibody (223H7) (10  $\mu$ g/mL) diluted in the washing buffer. Mix well and incubate for 30 minutes at room temperature.
- 6) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 7) Add 30  $\mu$ L of 1:50 Biotin labeled anti-mouse I-A<sup>b</sup> (A<sup>a</sup> $\beta$ <sup>b</sup>) (BD Biosciences; code no. 06042D) diluted with the washing buffer. Mix well and incubate for 15 minutes at room temperature.
- 8) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 9) Add 30  $\mu$ L of 1:50 PE labeled streptavidin (MBL; code no. IM-0557) diluted with the washing buffer. Mix well and incubate for 15 minutes at room temperature.
- 10) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 11) Resuspend the cells with 500  $\mu$ L of the washing buffer and analyze by a flow cytometer.

(Positive control for Flow cytometry; mouse splenocyte)