# For Research Use Only. Not for use in diagnostic procedures.



#### MONOCLONAL ANTIBODY

## Anti-His-tag mAb-Alexa Fluor® 488

Code No.CloneSubclassQuantityConcentrationD291-A48OGHisMouse IgG2a κ50 μL1 mg/mL

**BACKGROUND:** The His-tag (6xHis-tag) is one of the most common tags used to facilitate the purification of recombinant proteins. Metal chelate affinity chromatography is widely used for purification of His-tagged proteins. This specific antibody is useful tool for monitoring of the His-tagged proteins, and recognizes His-tags placed at N-terminal, C-terminal, and internal regions of the recombinant proteins.

**SOURCE:** This antibody was purified from hybridoma (clone OGHis) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell SP-1 with Balb/c mouse splenocyte immunized with 6xHis-tagged protein.

**FORMULATION:** 50 µg of IgG in 50 µL volume of PBS containing 1% BSA and 0.1% ProClin 150.

**STORAGE:** This antibody solution is stable for one year from the date of purchase when stored at 4°C.

**REACTIVITY:** This antibody recognizes His-tagged protein on Immunocytochemistry and Flow cytometry.

#### **APPLICATIONS:**

Immunocytochemistry; 1 μg/mL

Flow cytometry; 0.5 µg/mL (final concentration)

\*Please refer to the data sheet (MBL; code no. D291-3) for other applications.

Detailed procedure is provided in the following **PROTOCOLS**.

#### **INTENDED USE:**

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#### **REFERENCES:**

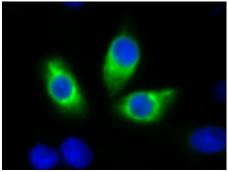
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The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

#### **PROTOCOLS:**

#### **Immunocytochemistry**

- 1) Culture the cells in the appropriate condition on a glass slide. (For example, spread 1 x 10<sup>4</sup> cells of transfectant cells for one slide, then incubate in a CO<sub>2</sub> incubator overnight.)
- 2) Wash the cells 3 times with PBS.
- 3) Fix the cells by immersing the slide in PBS containing 4% paraformaldehyde for 10 minutes at room temperature.
- 4) Wash the glass slide 3 times with PBS.
- 5) Immerse the slide in PBS containing 0.2% Triton X-100 for 10 minutes at room temperature.
- 6) Wash the glass slide 3 times with PBS.
- 7) Add the primary antibody diluted with PBS containing 2% FCS as suggested in the **APPLICATIONS** onto the cells and incubate for 30 minutes at room temperature. (Optimization of antibody concentration or incubation condition is recommended if necessary.)
- 8) Wash the glass slide twice with PBS.
- 9) Counter stain with DAPI for 5 minutes at room temperature.
- 10) Wash the glass slide twice with PBS.
- 11) Wipe excess buffer off the slide but take care not to touch the cells. Never leave the cells to dry.
- 12) Promptly add mounting medium onto the slide, then put a cover slip on it.



Immunocytochemical detection of His-tagged calnexin expressed in HeLa using D291-A48.

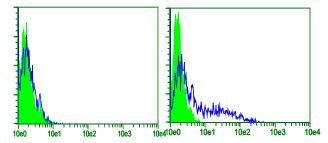
Green: Anti-His-tag mAb-Alexa Fluor® 488 (MBL; code no. D291-A48)

Blue: DAPI

### Flow cytometric analysis for floating cells

We usually use Fisher tubes or equivalents as reaction tubes for all steps described below.

- 1) Wash the cells 3 times with washing buffer [PBS containing 2% fetal calf serum (FCS) and 0.09% NaN<sub>3</sub>].
  - \*Azide may react with copper or lead in plumbing system to form explosive metal azides. Therefore, always flush plenty of water when disposing materials containing azide into drain.
- 2) Add 100  $\mu$ L of 4% paraformaldehyde (PFA) to the cell pellet after tapping. Mix well, then fix the cells for 10 minutes at room temperature.
- 3) Wash the cells twice with washing buffer.
- 4) Add 100  $\mu$ L of PBS containing 0.2% Triton X-100 to the cell pellet after tapping. Mix well, then permeabilize the cells for 10 minutes at room temperature.
- 5) Wash the cells once with washing buffer.
- 6) Add the primary antibody diluted with washing buffer as suggested in the **APPLICATIONS** onto the cells and incubate for 30 minutes at room temperature. (Optimization of antibody concentration or incubation condition is recommended if necessary.)
- 7) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration.
- 8) Resuspend the cells with 500  $\mu L$  of the washing buffer and analyze by a flow cytometer.



Flow cytometric analysis of HeLa (left) and His-tagged GM130 expressed in HeLa (right). Shaded histograms indicate the reaction of isotypic control to the cells. Open histograms indicate the reaction of D291-A48 to the cells.

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