

MONOCLONAL ANTIBODY

Anti-His-tag mAb-Agarose

Code No.	Clone	Subclass	Quantity
D291-8	OGHis	Mouse IgG2a κ	Gel: 200 μ L

BACKGROUND: The His-tag (6xHis-tag) is one of the most common tags used to facilitate the purification of recombinant proteins in *Escherichia coli* or other expression systems.

This product is useful tool for immunoprecipitation of the His-tagged proteins, and it recognizes His-tags placed at N-terminal, C-terminal, and internal regions of the recombinant proteins.

SOURCE: This antibody was purified from hybridoma (clone OGHis) supernatant using protein A agarose. This hybridoma was established by fusion of mouse myeloma cell SP-1 with Balb/c mouse splenocyte immunized with 6xHis tagged protein.

FORMULATION: 200 μ g of anti-His-tag monoclonal antibody covalently coupled to 200 μ L of agarose gel and provided as a 50% gel slurry suspended in PBS containing preservative (0.1% ProClin 150) for a total volume of 400 μ L.

STORAGE: This antibody solution is stable for one year from the date of purchase when stored at 4°C.

REACTIVITY: This antibody recognizes His-tag peptide sequence on Immunoprecipitation.

APPLICATION:

Immunoprecipitation: 20 μ L of gel slurry/300 μ L of cell extract from 1 x 10⁶ cells

Detailed procedure is provided in the following **PROTOCOL**.

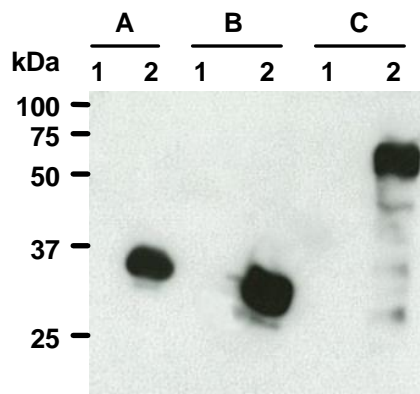
INTENDED USE:

For Research Use Only. Not for use in diagnostic procedures.

REFERENCES:

- 1) Kawada, J., *et al.*, *Int. J. Cancer* **130**, 584-592 (2012)
- 2) Ueyama, T., *et al.*, *J. Biol. Chem.* **286**, 40693-40705 (2011)
- 3) Suzuki, T., *et al.*, *Biochem. Biophys. Res. Commun.* **409**, 70-74 (2011)
- 4) Hiragami-Hamada, K., *et al.*, *Mol. Cell Biol.* **31**, 1186-1200 (2011)

Clone OGHis is used in these references.



Immunoprecipitation of His-tag from N-terminal His-tagged protein (A), Internal His-tagged protein (B), C-terminal His-tagged protein (C) with agarose conjugated mouse IgG1 isotype control, M075-8 (1) or D291-8 (2). After immunoprecipitated with the antibody, immunocomplexes were resolved on SDS-PAGE and immunoblotted with D291-7.

The descriptions of the following protocols are examples. Each user should determine the appropriate condition.

PROTOCOL:

Immunoprecipitation

- 1) Wash the cells 3 times with PBS and suspend with 10 volume of cold Lysis buffer (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.05% NP-40) containing appropriate protease inhibitors. Incubate it at 4°C with rotating for 30 minutes, then sonicate briefly (up to 10 seconds).
- 2) Centrifuge the tube at 12,000 x g for 10 minutes at 4°C and transfer the supernatant to another tube.
- 3) Add agarose gel as suggested in the **APPLICATIONS** into 300 μ L of the supernatant. Mix well and incubate with gentle agitation for 30-120 minutes at 4°C.
- 4) Wash the beads 3-5 times with the cold Lysis buffer (centrifuge the tube at 2,500 x g for 10 seconds).
- 5) Resuspend the agarose in 20 μ L of Laemmli's sample buffer, boil for 3-5 minutes, and centrifuge for 5 minutes.
- 6) Load 5 μ L of the sample per lane in a 1-mm-thick SDS-polyacrylamide gel and carry out electrophoresis.
- 7) Blot the protein to a polyvinylidene difluoride (PVDF) membrane at 1 mA/cm² for 1 hour in a semi-dry transfer system (Transfer Buffer: 25 mM Tris, 190 mM glycine, 20% MeOH). See the manufacturer's manual for precise transfer procedure.

- 8) To reduce nonspecific binding, soak the membrane in 10% skimmed milk (in PBS, pH 7.2) for 1 hour at room temperature, or overnight at 4°C.
- 9) Incubate the membrane with 1:5,000 of Anti-His-tag mAb-HRP-DirecT (MBL; code no. D291-7) diluted with PBS, pH 7.2 containing 1% skimmed milk for 1 hour at room temperature. (The concentration of antibody to be used will be depended on the condition.)
- 10) Wash the membrane with PBS-T [0.05% Tween-20 in PBS] (5 minutes x 3 times).
- 11) Wipe excess buffer on the membrane, then incubate it with appropriate chemiluminescence reagent for 1 minute. Remove extra reagent from the membrane by dabbing with paper towel, and seal it in plastic wrap.
- 12) Expose to an X-ray film in a dark room for 30 seconds. Develop the film as usual. The condition for exposure and development may vary.

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